

## 1 **Supplementary Materials for**

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## 3 **Lysosomal processing alters the specificity of sulfatide analogues for NKT cells and** 4 **subsequent immune responses in cancer**

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## 6 **Supplementary Methods**

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### 8 **General experimental for synthesis of sulfatide analogues**

9 Tetrahydrofuran (THF) was dried using a solvent dispensing system (SDS) with a column of  
10 neutral alumina. Pyridine, toluene, dimethylformamide (DMF), methylene chloride (CH<sub>2</sub>Cl<sub>2</sub>),  
11 deuterated chloroform (CDCl<sub>3</sub>), methanol (MeOH), deuterated methanol (CD<sub>3</sub>OD) and ethanol  
12 (EtOH) were dried over 4Å molecular sieves (MS). The other reagents were purchased from  
13 Acros, Alfa Aesar, Oakwood or Aldrich and used without further purification.

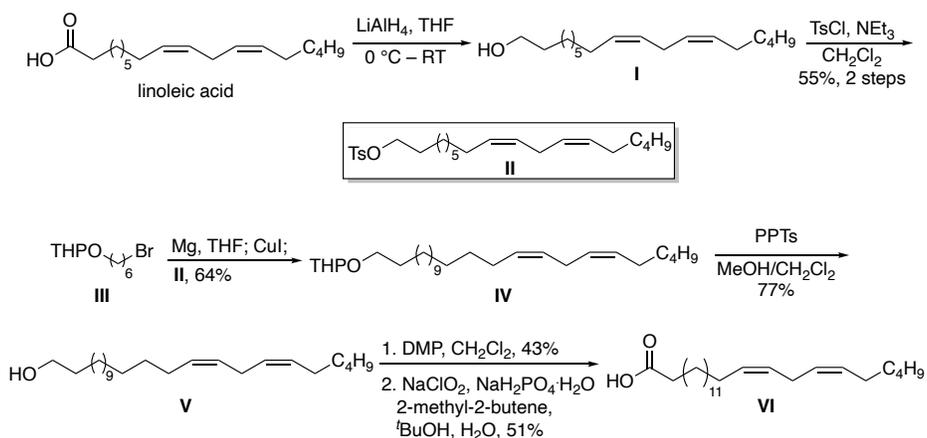
14 All reactions were conducted under an atmosphere of N<sub>2</sub> in glassware that had been dried  
15 overnight in an oven at 120 °C. Where appropriate, control of the reaction temperature was  
16 achieved with a solid CO<sub>2</sub>/acetone bath, an ice bath or a heated oil bath.

17 <sup>1</sup>H NMR spectra were recorded at 500 MHz or 400 MHz, and chemical shifts are calibrated to  
18 the residual CHCl<sub>3</sub> peak in CDCl<sub>3</sub> at 7.26 ppm, to the TMS peak at 0.0, or to the residual CD<sub>3</sub>OH  
19 peak in CD<sub>3</sub>OD at 3.34 ppm. <sup>13</sup>C NMR spectra were recorded at 125 MHz or 100 MHz and  
20 calibrated to the residual CHCl<sub>3</sub> peak in CDCl<sub>3</sub> at 77.23 or to the residual CD<sub>3</sub>OH peak in  
21 CD<sub>3</sub>OD at 49.5 ppm. The following abbreviations are used for peak multiplicities: app  
22 (apparent), s (singlet); br s (broadened singlet); d (doublet); dd (doublet of doublets); ddd

23 (doublet of doublet of doublets); dt (doublet of triplets); tt (triplet of triplets) t (triplet); q  
 24 (quartet); quin (quintet); m (multiplet). Coupling constants,  $J$ , are reported in Hertz (Hz).  
 25 IR spectra were recorded on a Bruker FT-IR spectrometer. High-resolution mass spectra  
 26 (HRMS) were obtained on an AccuTOF instrument equipped with a DART ionization source.  
 27 Melting points were observed in open Pyrex capillary tubes and are uncorrected. Specific  
 28 rotations  $[\alpha]_D$  were obtained on a JASCO polarimeter using the sodium D-line as a source, and  
 29 the concentration (c) is expressed in g per 100 mL.  
 30 Flash chromatography was performed on Silica Gel, 40 micron, 32-63 flash silica from Sorbent.  
 31 Thin layer chromatography was performed on silica gel (Silicycle Silica Gel 60 F<sub>254</sub> glass plates).  
 32 Compounds were visualized by UV, 5% phosphomolybdic acid in ethanol, 0.5% potassium  
 33 permanganate in water or a solution of ethanol/H<sub>2</sub>SO<sub>4</sub>/AcOH/*p*-anisaldehyde (135:5:1.5:3.7).  
 34 Ceric molybdate in a solution of H<sub>2</sub>O/ammonium molybdate/ceric ammonium molybdate/ H<sub>2</sub>SO<sub>4</sub>  
 35 (235 mL: 12 g: 0.5 g: 15mL) was used for sulfatides.

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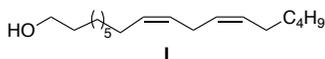
### 37 Preparation of 15Z,18Z-Tetracosadienoic acid (VI)



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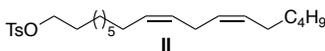
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42 **9Z,12Z-Octadecadien-1-ol (I).** Linoleic acid (1.0 g, 3.5 mmol) was dissolved in dry THF (40  
 43 mL) under N<sub>2</sub>, and the solution was cooled to 0 °C. After 10 min, LiAlH<sub>4</sub> (2M in THF, 5.3 mL,  
 44 10.6 mmol) was added dropwise over 2 min. The solution was stirred at 0 °C for 1 h then was  
 45 allowed to warm to rt over 2.5 h. The reaction mixture was then cooled to 0 °C, and the excess  
 46 LiAlH<sub>4</sub> was carefully quenched with saturated aqueous NH<sub>4</sub>Cl (20 mL). The organic layer was  
 47 separated, and the aqueous layer was extracted with EtOAc (3 x 30 mL). The combined organic  
 48 extracts were dried (MgSO<sub>4</sub>) and concentrated. The crude colorless oil (**I**) was moved forward  
 49 without purification. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 5.41–5.30 (m, 4H), 3.64–3.60 (m, 2H), 2.78  
 50 (t, *J* = 6.3 Hz, 2H), 2.13–2.10 (brs, 1H), 2.08–2.03 (m, 4H), 1.59–1.53 (m, 2H), 1.40–1.29 (m,  
 51 16 H), 0.90 (t, *J* = 6.0 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 130.2, 130.1, 128.0, 127.9, 62.8,  
 52 32.8, 31.5, 29.7, 29.5, 29.4, 29.3, 29.2, 27.2, 27.2, 25.8, 25.6, 22.6, 14.0.

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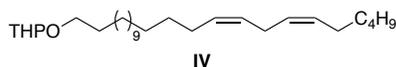


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55 **9Z,12Z-Octadecadiene tosylate (II).** Triethyl amine (0.60 mL, 4.3 mmol) and DMAP (48 mg,  
 56 0.40 mmol) were added to 9Z,12Z-octadecadien-1-ol (**I**) (1.0 g, 3.9 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (3.8  
 57 mL) at 0 °C. After 10 min, TsCl (0.78 g, 4.1 mmol) was added, and the solution was allowed to  
 58 warm to rt overnight. The reaction was diluted with CH<sub>2</sub>Cl<sub>2</sub> (15 mL) and washed with saturated  
 59 aqueous NH<sub>4</sub>Cl (25 mL). The organic layer was separated, and the aqueous layer was extracted  
 60 with CH<sub>2</sub>Cl<sub>2</sub> (3 x 25 mL). The combined organic extracts were dried (MgSO<sub>4</sub>) and concentrated.  
 61 Purification via flash column chromatography (hexanes/EtOAc 95:5) yielded **II** as a colorless oil  
 62 (0.92 g, 55% over two steps): IR (neat) 2927, 2857, 1357, 1174 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz,  
 63 CDCl<sub>3</sub>) δ 7.80 (d, *J* = 8.2 Hz, 2H), 7.35 (d, *J* = 8.1 Hz, 2H), 5.43–5.31 (m, 4H), 4.04 (t, *J* = 6.5

64 Hz, 2H), 2.79 (t,  $J = 6.2$  Hz, 2H), 2.46 (s, 3H), 2.09–2.03 (m, 4 H), 1.68–1.61 (m, 2H), 1.41–1.23  
 65 (m, 16H), 0.89 (t,  $J = 6.7$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  144.6, 133.3, 130.2, 130.0,  
 66 129.8, 128.1, 127.9, 127.8, 70.6, 31.5, 29.6, 29.4, 29.3, 29.1, 28.9, 28.8, 27.2, 27.2, 25.6, 25.3,  
 67 22.6, 21.6, 14.1.

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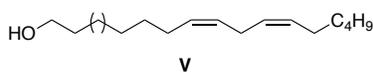


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70 **1-(2-Tetrahydro-2H-pyranyl)oxy-15Z,18Z-tetracosadiene (IV)**. Magnesium turnings (0.11 g,  
 71 5.0 mmol) were added to a flame dried 3-neck round bottom equipped with a reflux condenser.  
 72 The flask was flame dried a second time before adding a crystal of  $\text{I}_2$  and dry THF (4.5 mL). 2-  
 73 [(6-Bromohexyl)oxy]tetrahydro-2H-pyran (**III**)(1) (1.2 g, 4.6 mmol), was added in two portions.  
 74 Approximately one third of **III** was added to the flask, which was then heated with a heat gun  
 75 until the solution turned colorless. Once the color disappeared, remaining **III** was added while  
 76 maintaining reflux with the heat gun. The round bottom was then placed in a 60 °C oil bath to  
 77 stir for 40 min. The Grignard reagent was then added dropwise over 5 min to a suspension of  $\text{CuI}$   
 78 (0.43 g, 2.3 mmol) in dry THF (7.5 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 40  
 79 min. 9Z,12Z-Octadecadiene tosylate (**II**) (0.48 g, 1.1 mmol) in dry THF (7.5 mL) was added  
 80 dropwise over 5 min, and the mixture was stirred for 5 h at 0 °C, then allowed to warm to rt.  
 81 Saturated aqueous  $\text{NH}_4\text{Cl}$  (25 mL) was added, and the aqueous layer was extracted with  $\text{Et}_2\text{O}$  (3  
 82 x 30 mL). The combined organic extracts were washed with brine (40 mL), dried ( $\text{MgSO}_4$ ) and  
 83 concentrated. Purification via flash column chromatography (Hexanes/ $\text{EtOAc}$  99:1) on silica gel  
 84 yielded **IV** as a pale yellow oil (0.35 g, 64%): IR (neat) 3009, 2922, 2852, 1033  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR  
 85 (400 MHz,  $\text{CDCl}_3$ )  $\delta$  5.42–5.31 (m, 4H), 4.60–4.58 (m, 1H), 3.88 (ddd,  $J = 10.9$ ,  $J = 7.4$ ,  $J = 2.8$   
 86 Hz, 1H), 3.75 (ddd,  $J = 9.6$ ,  $J = 6.9$ ,  $J = 6.9$  Hz, 1H), 3.54–3.48 (m, 1H), 3.40 (ddd,  $J = 9.5$ ,  $J =$

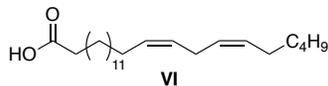
87 6.6,  $J = 6.6$  Hz, 1H), 2.79 (t,  $J = 6.4$  Hz, 2H), 2.09–2.04 (m, 4H), 1.89–1.81 (m, 1H), 1.76–1.70  
 88 (m, 1H), 1.64–1.50 (m, 6H), 1.41–1.29 (m, 28H), 0.89 (t,  $J = 6.5$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  
 89  $\text{CDCl}_3$ )  $\delta$  130.1, 128.0, 127.9, 98.8, 67.7, 62.2, 31.5, 30.8, 29.8, 29.7, 29.6, 29.6, 29.5, 29.4, 29.3,  
 90 27.2, 27.2, 26.3, 25.6, 25.5, 22.6, 19.7, 14.0; HRMS (ESI) calcd for  $\text{C}_{29}\text{H}_{55}\text{O}_2$   $[\text{M} + \text{H}]^+$   $m/z$   
 91 435.4202, found 435.4169.

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 93



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 95 **15Z,18Z-Tetracosadien-1-ol (V).** 1-(2-Tetrahydro-2H-pyranyl)oxy-15Z,18Z-tetracosadiene (**IV**)  
 96 (0.35 g, 0.81 mmol) was dissolved in  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  (1:1, 3.8 mL) followed by the addition of  
 97 PPTS (20 mg, 0.081 mmol). The reaction was stirred at 45 °C for 8 h. The MeOH was  
 98 evaporated, and the residue was diluted with  $\text{CH}_2\text{Cl}_2$  (20 mL) and  $\text{H}_2\text{O}$  (20 mL). The organic  
 99 layer was separated, and the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$  (3 x 10 mL). The  
 100 combined organic extracts were dried ( $\text{MgSO}_4$ ) and concentrated. Purification via flash column  
 101 chromatography (hexanes/EtOAc 95:5) yielded **V** as a pale yellow oil (0.22 g, 77%): IR (neat)  
 102 3400 (br), 2917, 2849, 1462, 1071, 683  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  5.41–5.30 (m, 4H),  
 103 3.64 (t,  $J = 6.6$  Hz, 2H), 2.77 (t,  $J = 6.5$  Hz, 2H), 1.57 (quin,  $J = 7.3$  Hz, 4H), 1.39–1.26 (m,  
 104 31H), 0.89 (t,  $J = 6.6$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  130.4, 128.2, 33.0, 31.8, 29.8,  
 105 29.8, 29.8, 29.7, 29.6, , 29.6, 27.5, 27.4, 26.0, 25.9, 14.3; HRMS (ESI) calcd for  $\text{C}_{24}\text{H}_{47}\text{O}$   $[\text{M} +$   
 106  $\text{H}]^+$   $m/z$  351.3621, found 351.3621.

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111 **15Z,18Z-Tetracosadienoic acid (VI).** Dess-Martin periodinane (0.16 g, 0.38 mmol) was added

112 to a solution of 15Z,18Z-tetracosadien-1-ol (V) (0.12 g, 0.35 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (1.3 mL) at 0

113 °C. The reaction mixture was stirred at rt for 6h. The reaction mixture was filtered through a pad

114 of celite, and the celite was washed with CH<sub>2</sub>Cl<sub>2</sub> (10 mL). The combined filtrates were

115 concentrated and purified by flash column chromatography on silica gel (hexanes/EtOAc, 90:10)

116 to provide 15Z,18Z-tetracosadienal as a colorless oil (53 mg, 43%): IR (neat) 2920, 2850, 1700,

117 1650, 1510, 1100, 1050 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.76 (s, 1H), 5.41–5.30 (m, 4H),

118 2.77 (t, *J* = 6.0 Hz, 2H), 2.41 (t, *J* = 7.2 Hz, 2H), 2.04 (m, 4H), 1.61 (m, 2H), 1.34–1.26 (m,

119 26H), 0.89 (t, *J* = 6.9 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 203.2, 130.4, 128.2, 44.1, 31.8,

120 29.9, 29.9, 29.8, 29.6, 29.6, 29.6, 29.4, 27.5, 27.4, 25.9, 22.8, 22.3, 14.3. NaH<sub>2</sub>PO<sub>4</sub> (0.14 g, 1.0

121 mmol) was added to a mixture of 15Z,18Z-tetracosadienal (0.060 g, 0.18 mmol) and 2-methyl-2-

122 butene (0.4 mL, 4 mmol) in *t*BuOH (7 mL) and H<sub>2</sub>O (1.5 mL) at 0 °C. NaClO<sub>2</sub> (0.020 g, 0.22

123 mmol) was added in small portions and the mixture stirred for 6 h. One more equiv of NaClO<sub>2</sub>

124 was added, and the reaction mixture was left in the fridge overnight. The next day, TLC still

125 showed remaining aldehyde; so another equiv of NaClO<sub>2</sub> was added, and the reaction mixture

126 was stirred for 40 min at 0 °C. After this, TLC showed complete consumption of the aldehyde.

127 Saturated aqueous Na<sub>2</sub>SO<sub>3</sub> and pH7 phosphate buffer (1:1, 2 mL) were added. The product was

128 extracted with EtOAc (3 X 10 mL). The combined organic extracts were washed with sat. NH<sub>4</sub>Cl

129 (5 mL) and brine (5 mL), dried (MgSO<sub>4</sub>), filtered and concentrated to give VI with ~10% of

130 inseparable *E/Z*-stereoisomers (0.036 g, 51%) as a colorless oil: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ

131 5.41–5.30 (m, 4H), 2.77 (t, *J* = 5.8 Hz, 2H), 2.34 (t, *J* = 7.5 Hz, 2H), 2.05 (m, 4H), 1.63 (quin, *J*

132 = 7.2 Hz, 2H), 1.40–1.26 (m, 26H), 0.91–0.86 (m, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 180.5,

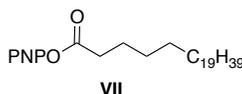
133 130.4, 128.2, 34.3, 31.8, 30.0, 29.9, 29.8, 29.7, 29.6, 29.6, 29.5, 29.3, 27.5, 27.4, 25.8, 24.9, 22.8,  
 134 14.3.

135

136 **General *p*-nitrophenyl ester preparation**

137 *p*-Nitrophenol (1.1 equiv) and DMAP (0.2 equiv.) were added to a flask charged with carboxylic  
 138 acid (1.0 equiv.) in dry CH<sub>2</sub>Cl<sub>2</sub> (0.014 M), and the solution was stirred for 15 min. DCC (1.04  
 139 equiv) in dry CH<sub>2</sub>Cl<sub>2</sub> (0.12 M) was then added slowly. The reaction mixture was allowed to stir  
 140 at rt overnight, then filtered through a pad of celite. The celite was washed with CH<sub>2</sub>Cl<sub>2</sub>, and the  
 141 filtrate was concentrated. Purification via flash chromatography on silica gel (petroleum  
 142 ether/EtOAc, 95:5) yielded PNP-activated esters **VII-IX**.

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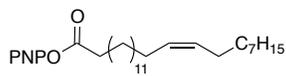


144

145 ***p*-Nitrophenyltetraacosanoate (VII)**. Compound **VII** was prepared from tetraacosanoic acid and  
 146 was isolated as a white solid (0.29 g, 73%): mp 81.9–82.2 °C; IR (neat) 2916, 2849, 1752, 1535,  
 147 1347, 1203, 1136, 1107, 927, 868, 717 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.29–8.25 (m, 2H),  
 148 7.29–7.26 (m, 2H), 2.59 (t, *J* = 7.4 Hz, 2H), 1.76 (quin, *J* = 7.3 Hz 2H), 1.45–1.26 (m, 40H),  
 149 0.88 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 171.5, 155.8, 145.4, 125.4, 122.6, 34.6,  
 150 32.2, 29.9, 29.8, 29.7, 29.6, 29.4, 29.3, 25.0, 22.9, 14.3; HRMS (ESI) calcd for C<sub>30</sub>H<sub>52</sub>NO<sub>4</sub> [M+  
 151 H]<sup>+</sup> *m/z* 490.3891, found 490.3921.

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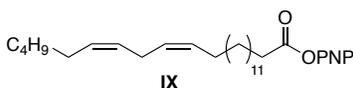


VIII

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155 ***p*-Nitrophenyl 15*Z*-tetracosenoate (VIII)**. Compound VIII was prepared from nervonic acid  
 156 and was isolated as a colorless solid (0.50 g, 73%): mp 35.5–36.0 °C; IR (neat) 2916, 2850,  
 157 1753, 1593, 1536, 1490, 1471, 1350, 1203, 1138, 926, 868, 717 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz,  
 158 CDCl<sub>3</sub>) δ 8.18 (d, *J* = 8.7 Hz, 2H), 7.19 (d, *J* = 8.6 Hz, 2H), 5.27 (m, 2H), 2.51 (t, *J* = 7.3 Hz,  
 159 2H), 1.96–1.91 (m, 4H), 1.68 (quin, *J* = 7.0 Hz, 2H), 1.34–1.19 (m, 32H), 0.80 (t, *J* = 6.7 Hz,  
 160 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 171.4, 155.7, 145.4, 130.1, 130.0, 125.3, 122.6, 34.5, 32.1,  
 161 30.0, 29.8, 29.8, 29.7, 29.6, 29.5, 29.4, 29.2, 27.4, 24.9, 22.9, 14.3; HRMS (ESI) calcd for  
 162 C<sub>30</sub>H<sub>50</sub>NO<sub>4</sub> [M + H]<sup>+</sup> *m/z* 488.3734, found 488.3755.

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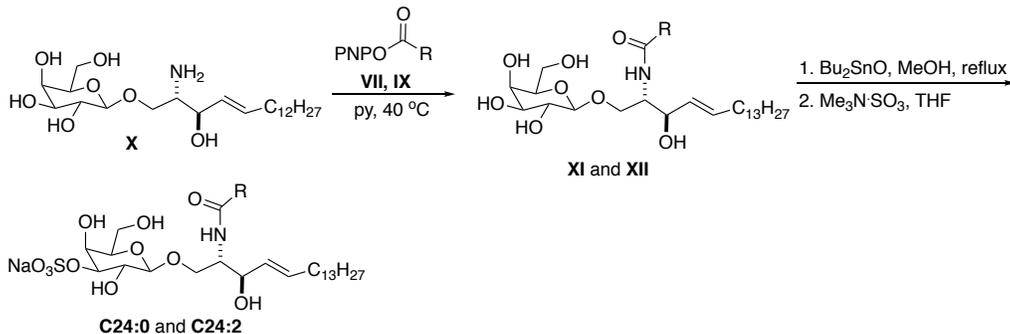
IX

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165 ***p*-Nitrophenyl 15*Z*,18*Z*-tetracosadienoate (IX)**. Compound IX was prepared from 15*Z*,18*Z*-  
 166 tetracosadienoic acid (VI) and was isolated as a low melting solid (29.0 mg, 45%): IR (neat)  
 167 2922, 2852, 1768, 1593, 1524, 1490, 1464, 1345, 1208, 1098, 863 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz,  
 168 CDCl<sub>3</sub>) δ 8.18 (d, *J* = 9.0 Hz, 2H), 7.19 (d, *J* = 8.9 Hz, 2H), 5.33–5.21 (m, 4H), 2.71–2.64 (m,  
 169 2H), 2.52 (t, *J* = 7.4 Hz, 2H), 1.97 (m, 4H), 1.68 (quin, *J* = 7.2 Hz, 2H), 1.36–1.20 (m, 26H),  
 170 0.81 (t, *J* = 6.1 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 171.4, 155.7, 145.4, 130.3, 128.1, 125.3,  
 171 122.6, 34.5, 31.7, 29.8, 29.8, 29.6, 29.5, 29.4, 29.2, 27.6, 27.4, 25.8, 24.9, 22.8, 14.2; HRMS  
 172 (ESI) calcd for C<sub>30</sub>H<sub>48</sub>NO<sub>4</sub> [M + H]<sup>+</sup> *m/z* 486.3578, found 486.3570.

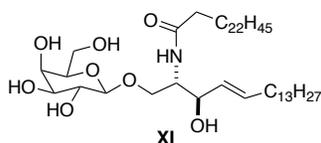
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175 **Preparation of sulfatides C24:0 and C24:2**

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179 **(2*S*,3*R*,4*E*)-1-(β-D-Galactopyranosyloxy)-2-(*N*-tetracosanoylamino)octadec-4-en-3-ol (XI).**180 *p*-Nitrophenyltetracosanoate (**VII**) (0.04 g, 0.08 mmol) was added to a solution of (2*S*,3*R*,4*E*)-181 2-amino-1-(β-galactopyranosyloxy)octadec-4-en-3-ol(**X**) (0.03 g, 0.7 mmol) in pyridine (1

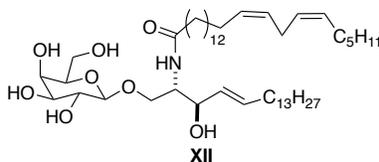
182 mL). The solution was stirred in a preheated oil bath at 40 °C overnight. The solution was

183 concentrated and purified by flash column chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 90:10)184 to give **XI** as a white solid (17.2 mg, 32%): mp 182.0–183.0 °C; [α]<sup>25</sup><sub>D</sub> 1.10 (c 1.25,185 CHCl<sub>3</sub>/MeOH); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 7.27 (d, *J* = 8.8 Hz, 1H), 5.69 (dt, *J*186 = 15.2, 7.2 Hz, 1H), 5.45 (dd, *J* = 15.0, 6.6 Hz, 1H), 4.21 (d, *J* = 6.8 Hz, 1H), 4.00 (br s, 1H),187 3.88 (br s, 1H), 3.81–3.72 (m, 2H), 3.61 (m, 1H), 3.56–3.50 (m, 3H), 2.77 (t, *J* = 6.9 Hz, 2H),188 2.02 (dt, *J* = 6.9, 6.9 Hz, 2H), 1.60–1.57 (m, 2H), 1.36–1.26 (m, 62H), 0.88 (t, *J* = 6.4 Hz, 6H);189 <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 175.5, 135.0, 130.2, 104.7, 76.0, 74.4, 73.0, 72.3,

190 70.0, 69.6, 62.5, 54.5, 50.1, 33.2, 32.7, 30.5, 30.5, 30.4, 30.3, 30.2, 30.2, 26.8, 23.5, 14,6; HRMS

191 (ESI) calcd for C<sub>48</sub>H<sub>94</sub>NO<sub>8</sub> [M + H]<sup>+</sup> *m/z* 812.6974, found 812.6982.

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193

194 **(2*S*,3*R*,4*E*)-1-(β-*D*-Galactopyranosyloxy)-2-(*N*-15*Z*,18*Z*-tetracosadienoylamino)octadec-4-**195 **en-3-ol (XII).** *p*-Nitrophenyl 15*Z*,18*Z*-tetracosadieneoate (**IX**) (28 mg, 0.06 mmol) was added to196 a solution of (2*S*,3*R*,4*E*)-2-amino-1-(β-galactopyranosyloxy)octadec-4-en-3-ol(**2**) (**X**) (25 mg,

197 0.60 mmol) in pyridine (1 mL). The mixture was stirred in a preheated oil bath at 40 °C

198 overnight. The reaction was concentrated and purified by flash column chromatography on silica

199 gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 90:10) to give **XII** (21 mg, 46%) as an off white solid: mp 129.0–130.0 °C;200 IR (neat) 3302, 2915, 1641, 1544, 1467, 1082 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ201 5.70 (dt, *J* = 14.6, 6.6 Hz, 1H), 5.46 (m, 1H), 5.41–5.29 (m, 4H), 4.21 (d, *J* = 7.4 Hz, 1H), 4.00202 (ddd, *J* = 7.3, 3.7, 3.7 Hz, 1H), 3.82 (app d, *J* = 2.6 Hz, 1H), 3.81 (dd, *J* = 11.5, 6.6 Hz, 1H),203 3.75 (dd, *J* = 11.5, 5.0 Hz 1H), 3.62 (dd, *J* = 10.3, 3.2 Hz, 1H), 3.57–3.47 (m, 3H), 2.77 (t, *J* =204 6.2 Hz, 2H), 2.17 (t, *J* = 7.4 Hz, 2H), 2.07–1.99 (m, 6H), 1.59 (quin, *J* = 7.1 Hz, 2H), 1.40–1.27205 (m, 53H), 0.88 (t, *J* = 6.9 Hz, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 175.6, 135.0,

206 131.0, 130.3, 128.9, 104.8, 76.1, 74.5, 73.1, 72.4, 70.0, 69.7, 62.5, 54.5, 37.4, 33.2, 32.8, 32.4,

207 30.5, 30.5, 30.4, 30.3, 30.3, 30.2, 28.1, 28.1, 26.8, 26.5, 23.5, 23.4, 14.7; HRMS (ESI) calcd for

208 C<sub>48</sub>H<sub>90</sub>NO<sub>8</sub> [M + H]<sup>+</sup> *m/z* 808.6661, found 808.6660.

209

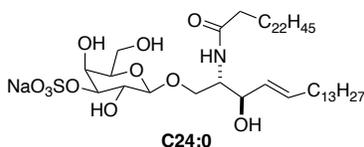
210 **General sulfation procedure**211 Glycolipids (1 equiv) and Bu<sub>2</sub>SnO (1.2 equiv) were refluxed in MeOH (0.016 M) for 2 h. The

212 solvent was evaporated under reduced pressure. The resulting dibutyl-stannylene complex was

213 treated with Me<sub>3</sub>N•SO<sub>3</sub> (2 equiv) in THF (2 mL)(3, 4). The mixture was stirred at rt from

214 between 2 and 6 h. TLC was used to monitor the reaction. The solvent was evaporated, and the  
 215 residue dissolved in a 1:1 mixture of CH<sub>2</sub>Cl<sub>2</sub>/MeOH (4 mL). Dowex (Na<sup>+</sup> resin) was added. The  
 216 mixture was then stirred for 10 min, followed by filtration and concentration. The crude product  
 217 was partitioned in a mixture of 1-butanol/H<sub>2</sub>O (1:1, v/v) and centrifuged. The supernatant (1-  
 218 butanol, containing the sulfatides) was collected and concentrated. Purification by flash column  
 219 chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 90:10 – 85:15) gave the sulfatides.

220



221

222 **(2*S*,3*R*,4*E*)-1-(3-*O*-Sodiumsulfonyl-β-*D*-galactopyranosyloxy)-2-(*N*-tetracosanoylamino)-**

223 **octadec-4-en-3-ol (C24:0)**. Following the general sulfation procedure, sulfatide **C24:0** was

224 isolated as a white solid (4.2 mg, 35%): mp 204.0–205.0 °C; [ $\alpha$ ]<sup>25</sup><sub>D</sub> 6.98 (*c* 0.38, CHCl<sub>3</sub>/MeOH,

225 3:2); IR (neat) 3400 (br), 2917, 2850, 1646, 1466, 1258, 1066 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz,

226 CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2)  $\delta$  7.71 (d, *J* = 8.8 Hz, 1H), 5.67 (dt, *J* = 15.3, 6.7 Hz, 1H), 5.42 (dd, *J* =

227 15.3, 6.6 Hz, 1H), 3.81–3.74 (m, 4H), 3.58–3.56 (m, 2H), 2.15 (t, *J* = 7.9 Hz, 2H), 2.00–1.98 (m,

228 2H), 1.66–1.45 (m, 2H), 1.41–1.25 (m, 62H), 0.87 (t, *J* = 6.9 Hz, 6H); <sup>13</sup>C NMR (100 MHz,

229 CHCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2)  $\delta$  175.7, 135.4, 130.3, 104.3, 81.4, 75.8, 72.7, 70.5, 69.7, 68.2, 62.2, 54.2,

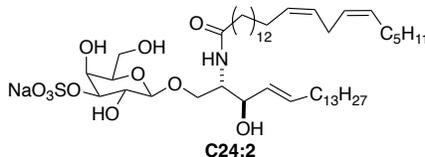
230 37.4, 33.4, 32.9, 31.5, 30.7, 30.6, 30.6, 30.5, 30.4, 30.3, 30.3, 26.9, 23.5, 14.8; HRMS (TOF) *m/z*

231 calcd for C<sub>48</sub>H<sub>92</sub>NO<sub>11</sub>S [M – Na]<sup>+</sup> 890.6397, found 890.6377.

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236 **(2*S*,3*R*,4*E*)-1-(3-*O*-Sodiumsulfonyl-β-*D*-galactopyranosyloxy)-2-(*N*-15*Z*,18*Z*-**237 **tetracosadienoylamino)octadec-4-en-3-ol (C24:2).** Sulfatide **C24:2** was isolated as an off

238 white solid (14.9 mg, 65%): mp 182.0–183.0 °C; IR (neat) 3370 (br), 2918, 2850, 1644, 1467,

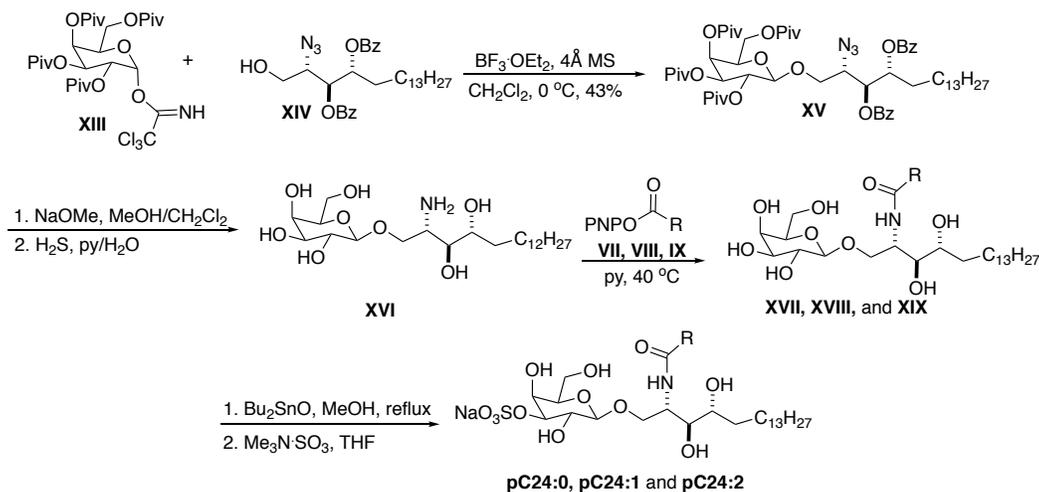
239 1258, 1066 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 5.70 (dt, *J* = 15.3, 6.6 Hz, 1H),240 5.44 (dd, *J* = 15.4, 7.4 Hz, 1H), 5.40–5.29 (m, 4H), 4.34 (d, *J* = 7.7 Hz, 1H), 3.64 (dd, *J* = 10.3,241 3.2 Hz, 1H), 3.57 (dd, *J* = 5.7, 5.7 Hz, 1H), 2.77 (d, *J* = 6.3 Hz, 2H), 2.17 (t, *J* = 7.6 Hz, 2H),242 2.08–2.00 (m, 6H), 1.65–1.51 (m, 2H), 1.40–1.27 (m, 48H), 0.91–0.86 (m, 6H); <sup>13</sup>C NMR (100243 MHz, CHCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 175.8, 135.2, 131.1, 130.4, 128.9, 104.5, 81.4, 75.8, 72.9, 70.6,

244 69.9, 68.6, 62.4, 54.5, 37.4, 33.2, 32.8, 32.4, 30.6, 30.5, 30.5, 30.4, 30.4, 30.3, 30.2, 28.1, 28.1,

245 26.9, 26.6, 23.5, 23.4, 14.7; HRMS (TOF) calcd for C<sub>48</sub>H<sub>92</sub>NO<sub>11</sub> [M – Na]<sup>+</sup> *m/z* 886.6078, found

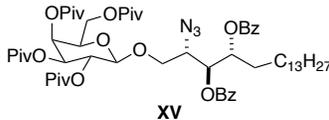
246 886.6058.

247

248 **Preparation of pC24:0, pC24:1 and pC24:2**

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250



251

252 **(2*S*,3*S*,4*R*)-2-Azido-3,4-dibenzoyloxy-1-(2,3,4,6-tetra-*O*-pivaloyl-β-*D*-galactopyrano-**253 **side)octadecane (XV).** (2,3,4,6-Tetra-*O*-pivaloyl-α-*D*-galactopyranoside)-1-trichloro-254 acetimidate(2) (**XIII**) (0.60 g, 0.99 mmol) and (2*S*,3*S*,4*R*)-2-azido-(3,4-dibenzoyloxy)octadecan-255 1-ol(5) (**XIV**) (0.45 g, 0.82 mmol) were dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> (13 mL), and the solution was

256 stirred in the presence of 4Å MS (600 mg) at rt for 10 min. The solution was then cooled to –10

257 °C. BF<sub>3</sub>•OEt<sub>2</sub> in dry CH<sub>2</sub>Cl<sub>2</sub> (1.46 μL in 2 mL) was added over 10 min; then the solution was

258 allowed to slowly warm to rt and stir for 1.5 h. The reaction mixture was diluted with petroleum

259 ether (50 mL) and then filtered. The filtrate was washed with saturated aqueous NaHCO<sub>3</sub> (10260 mL). The organic layer was separated, and the aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 X 15261 mL). The combined organic extracts were dried (MgSO<sub>4</sub>), filtered and concentrated. Purification262 by flash column chromatography on silica gel (petroleum ether/ EtOAc 95:5) gave **XV** (0.34 g,263 39%) as a colorless oil: [α]<sup>25</sup><sub>D</sub> –3.65 (*c* 1.00, CH<sub>2</sub>Cl<sub>2</sub>); IR (neat) 2926, 2103, 1728, 1480, 1261,264 1140, 710 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.00 (d, *J* = 7.8 Hz, 4H), 7.57 (m, 2H), 7.44 (m,265 4H), 5.49– 5.44 (m, 2H), 5.37 (d, *J* = 3.1 Hz, 1H), 5.22 (dd, *J* = 10.5, 8.1 Hz, 1H), 5.05 (dd, *J* =266 10.4, 3.2 Hz, 1H), 4.56 (d, *J* = 7.9 Hz, 1H), 4.08–4.02 (m, 2H), 3.98–3.90 (m, 4H), 1.88–1.80

267 (m, 2H), 1.43–1.32 (m, 3H), 1.29–1.20 (m, 30H), 1.14 (s, 9H), 1.09 (s, 9H), 1.08 (s, 9H), 0.86 (t,

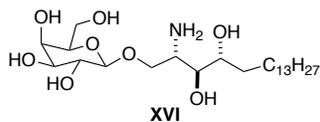
268 *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 177.9, 177.4, 177.0, 176.5, 133.7, 133.4, 130.0,

269 129.9, 129.8, 129.5, 128.7, 128.6, 100.9, 73.0, 71.2, 71.1, 68.7, 68.6, 66.7, 61.4, 61.1, 39.2, 38.9,

270 38.8, 32.0, 30.3, 29.8, 29.8, 29.8, 29.8, 29.8, 29.7, 29.7, 29.6, 29.5, 29.5, 29.5, 25.4, 22.8, 14.2;

271 HRMS (ESI) calcd for C<sub>58</sub>H<sub>88</sub>N<sub>3</sub>O<sub>14</sub> [M + H]<sup>+</sup> *m/z* 1050.6261, found 1050.6300.

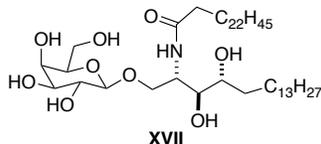
272



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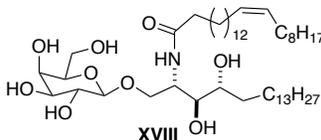
274 **(2*S*,3*S*,4*R*)-2-Amino-1-(β-*D*-galactopyranosyloxy)octadecan-3,4-diol (XVI).** NaOMe in  
 275 MeOH (0.50 M, 4.0 mL, 2.0 mmol) was added to a solution of (2*S*,3*S*,4*R*)-2-azido-3,4-  
 276 dibenzoyloxy-1-(2,3,4,6-tetra-*O*-pivaloyl-β-galactopyranoside)octadecane (**XV**) (296 mg, 0.28  
 277 mmol) in a mixture of CH<sub>2</sub>Cl<sub>2</sub>/MeOH (3.4/3.4 mL)(2). The solution was stirred at rt for 1.5 h.  
 278 The solution was then acidified with dowex (H<sup>+</sup>) resin. The mixture was filtered through a pad of  
 279 celite, and the celite was washed with a 1:1 mixture of CHCl<sub>3</sub> and MeOH (15 mL). The filtrate  
 280 was concentrated and triturated with petroleum ether/EtOAc (85:15) to give (2*S*,3*S*,4*R*)-2-azido-  
 281 1-(β-galactopyranosyloxy)octadecan-3,4-diol (134 mg, 94%) as a white solid: [α]<sup>25</sup><sub>D</sub> 18.9 (*c* 6.64,  
 282 CHCl<sub>3</sub>/MeOH, 3:2); IR (neat) 3355 (br), 2915, 2849, 2096, 1255, 1071 cm<sup>-1</sup>; <sup>1</sup>H NMR (400  
 283 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 4.28 (d, *J* = 7.2 Hz, 1H), 4.13 (dd, *J* = 10.6, 5.0 Hz, 1H), 3.96 (app  
 284 d, *J* = 10.3 Hz, 1H), 3.97 (s, 1H), 3.82 (dd, *J* = 11.5, 6.5 Hz, 1H), 3.70–3.63 (m, 4H), 3.58–3.49  
 285 (m, 3H), 1.67–1.56 (m, 2H), 1.42–1.25 (m, 24H), 0.87 (t, *J* = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz,  
 286 CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 104.3, 76.1, 74.9, 74.4, 72.7, 72.1, 69.9, 69.4, 63.1, 62.3, 33.2, 32.8, 30.6,  
 287 30.5, 30.5, 30.2, 26.6, 23.5, 14.7; HRMS (ESI) calcd for C<sub>24</sub>H<sub>48</sub>N<sub>3</sub>O<sub>8</sub> [M + H]<sup>+</sup> *m/z* 506.3436,  
 288 found 506.3511. The product was carried forward to reduction of the azide. A solution of  
 289 (2*S*,3*S*,4*R*)-2-azido-1-(β-galactopyranosyloxy)octadecan-3,4-diol (13 mg, 0.27 mmol) in a  
 290 mixture of pyridine/H<sub>2</sub>O (1:1, 7.6 mL) was saturated with H<sub>2</sub>S. The solution was stirred for 48  
 291 h(2). The solvent was evaporated to give **XVI** (136 mg, crude) as a yellowish brown powder,  
 292 which was carried forward without purification.

293



294  
 295 **(2*S*,3*S*,4*R*)-1-(β-D-Galactopyranosyloxy)-2-(*N*-tetracosanoylamino)octadecane-3,4-diol**  
 296 **(XVII)**. *p*-Nitrophenyltetracosanoate (**VII**) (30 mg, 0.07 mmol) was added to a solution of  
 297 (2*S*,3*S*,4*R*)-2-amino-1-(β-galactopyranosyloxy)octadecane-3,4-diol (**XVI**) (30 mg, 0.06 mmol) in  
 298 pyridine (1 mL). The mixture was stirred in a preheated oil bath at 40 °C overnight. The reaction  
 299 was concentrated and purified by flash column chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH,  
 300 95:5) to give **XVII** as a white solid (16 mg, 35%): mp 198.7–199.8 °C; [α]<sup>25</sup><sub>D</sub> 10.20 (*c* 0.49,  
 301 CHCl<sub>3</sub>/MeOH, 3:2); IR (neat) 3304, 2915, 2849, 1625, 1468, 1077, 718 cm<sup>-1</sup>; <sup>1</sup>H NMR (400  
 302 MHz, CDCl<sub>3</sub>/MeOD, 3:2) δ 4.25–4.12 (m, 2H), 3.87–3.86 (m, 1H), 3.82 (dd, *J* = 11.6, 6.7 Hz,  
 303 2H), 3.75–3.69 (m, 2H), 3.61–3.47 (m, 5H), 2.20 (t, *J* = 7.5 Hz, 2H), 1.68–1.52 (m, 4H), 1.44–  
 304 1.27 (m, 64H), 0.88 (t, *J* = 6.5 Hz, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>/MeOD, 3:2) δ 175.6, 104.8,  
 305 76.3, 75.4, 74.5, 73.3, 72.4, 70.2, 70.1, 51.5, 37.4, 33.2, 32.8, 31.3, 30.7, 30.6, 30.6, 30.5, 30.3,  
 306 30.3, 30.2, 26.8, 26.8, 23.5, 14.7; HRMS (TOF) *m/z* calcd for C<sub>48</sub>H<sub>96</sub>NO<sub>9</sub> [M + H]<sup>+</sup> 830.7080,  
 307 found 830.7052.

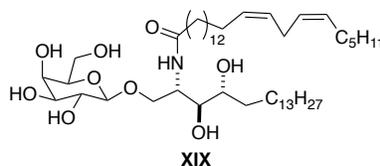
308



309  
 310 **(2*S*,3*S*,4*R*)-1-(β-D-Galactopyranosyloxy)-2-(*N*-15*Z*-tetracosenoylamino)octadecane-3,4-diol**  
 311 **(XVIII)**. *p*-Nitrophenyl 15*Z*-tetracosenoate (**VIII**) (33 mg, 0.07 mmol) was added to a solution  
 312 of (2*S*,3*S*,4*R*)-2-amino-1-(β-galactopyranosyloxy)octadecane-3,4-diol (**XVI**) (30 mg, 0.06 mmol)  
 313 in pyridine (1 mL). The mixture was stirred in a preheated oil bath at 40 °C overnight. The

314 reaction was concentrated and purified by flash column chromatography on silica gel  
 315 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 95:5) to give **XVIII** (19 mg, 38%) as a white solid: mp 169.0–171.0 °C; [α]<sup>25</sup><sub>D</sub>  
 316 7.38 (c 0.82, CHCl<sub>3</sub>/MeOH, 3:2); IR (neat) 3330, 2917, 2849, 1637, 1545, 1465, 1081, 1049, cm<sup>-1</sup>;  
 317 <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 5.38–5.30 (m, 2H), 4.24–4.20 (m, 2H), 4.16 (dd, *J*  
 318 = 10.3, 4.6 Hz, 1H), 3.87 (app d, *J* = 2.7 Hz, 1H), 3.82 (dd, *J* = 11.6, 6.8 Hz, 1H), 3.74 (ddd, *J* =  
 319 11.1, 4.8 Hz, 1H), 3.71 (dd, *J* = 10.2, 3.8 Hz, 1H), 3.61–3.47 (m, 5H), 2.20 (t, *J* = 7.6 Hz, 2H),  
 320 2.04–1.99 (m, 4H), 1.68–1.57 (m, 3H), 1.53–1.51 (m, 1H), 1.47–1.27 (m, 58H), 0.88 (t, *J* = 6.0  
 321 Hz, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 175.5, 130.7, 104.8, 76.2, 75.5, 74.5, 73.3,  
 322 72.4, 70.1, 70.1, 62.6, 51.5, 37.4, 33.6, 32.7, 30.6, 30.6, 30.5, 30.4, 30.3, 30.3, 30.2, 30.1, 28.0,  
 323 26.8, 26.7, 23.5, 14.6; HRMS (TOF) *m/z* calcd for C<sub>48</sub>H<sub>94</sub>NO<sub>9</sub> [M – H]<sup>+</sup> 828.6923, found  
 324 828.6915.

325

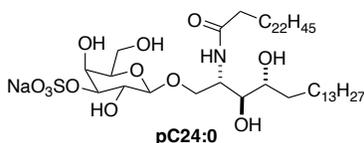


326

327 **(2*S*,3*S*,4*E*)-1-(β-*D*-Galactopyranosyloxy)-2-(*N*-15*Z*,18*Z*-tetracosadienoylamino)octadecan-**  
 328 **3,4-diol (XIX).** *p*-Nitrophenyl 15*Z*,18*Z*-tetracosadieneoate (**IX**) (28 mg, 0.06 mmol) was added  
 329 to a solution of (2*S*,3*S*,4*R*)-2-amino-1-(β-galactopyranosyloxy)octadecan-3,4-diol (**XVI**) (27.0  
 330 mg, 0.06 mmol) in pyridine (1 mL). The mixture was stirred in a preheated oil bath at 40 °C  
 331 overnight. The reaction was concentrated and purified by flash column chromatography on silica  
 332 gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 95:5) to give **XIX** (19 mg, 40%) as a white solid: mp 169.0–171.0 °C; [α]<sup>25</sup><sub>D</sub>  
 333 8.81 (c 1.86, CHCl<sub>3</sub>/MeOH, 3:2); IR (neat) 3302, 2918, 2850, 1637, 1467, 1082 cm<sup>-1</sup>; <sup>1</sup>H NMR  
 334 (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 5.41–5.29(m, 4H), 4.23–4.22 (m, 2H), 4.14–4.12 (m, 1H),

335 3.87 (app d,  $J = 2.0$  Hz, 1H), 3.82 (dd,  $J = 11.6, 6.8$  Hz, 1H), 3.75–3.69 (m, 2H), 3.61–3.47 (m,  
 336 5H), 2.77 (t,  $J = 6.1$  Hz, 2H), 2.20 (t,  $J = 7.4$  Hz, 2H), 2.07–2.01 (m, 4H), 1.68–1.51 (m, 5H),  
 337 1.44–1.27 (m, 51H), 0.88 (t,  $J = 4.8$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$   $\text{CDCl}_3/\text{CD}_3\text{OD}$ , 3:2)  
 338  $\delta$  175.6, 131.0, 128.9, 76.3, 75.5, 74.5, 73.3, 72.4, 70.1, 70.1, 62.6, 51.5, 37.4, 33.2, 32.8, 32.4,  
 339 30.7, 30.6, 30.4, 30.3, 30.3, 30.2, 28.1, 28.1, 26.8, 26.8, 26.5, 23.5, 23.4, 14.7; HRMS (TOF)  $m/z$   
 340 calcd for  $\text{C}_{48}\text{H}_{92}\text{NO}_9$   $[\text{M} - \text{H}]^+$  826.6767, found 826.6777.

341

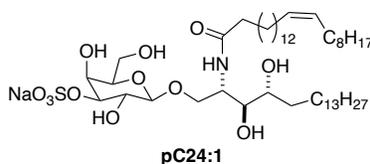


342

343 **(2*S*,3*S*,4*R*)-1-(3-*O*-sodiumsulfonyl- $\beta$ -D-galactopyranosyloxy)-2-(*N*-tetracosanoylamino)-**  
 344 **octadecane-3,4-diol (pC24:0).** The general sulfation procedure was followed, and sulfatide  
 345 **pC24:0** was isolated as a white solid (3.2 mg, 18%): mp 184.0–185.0 °C;  $[\alpha]_D^{25}$  10.43 ( $c$  0.49,  
 346  $\text{CHCl}_3/\text{MeOH}$ , 3:2); IR (neat) 3429 (br), 2917, 2850, 1632, 1467, 1224, 1070, 801  $\text{cm}^{-1}$ ;  $^1\text{H}$   
 347 NMR (400 MHz,  $\text{CDCl}_3/\text{CD}_3\text{OD}$ , 3:2)  $\delta$  5.35–5.32 (m, 1H), 3.81 (dd,  $J = 11.8, 7.2$  Hz, 1H),  
 348 3.76–3.71 (m, 2H), 3.66–3.64 (m, 1H), 3.59–3.56 (m, 1H), 2.15 (t,  $J = 7.5$  Hz, 2H), 2.04–2.00  
 349 (m, 1H), 1.60–1.51 (m, 4H), 1.41–1.26 (m, 62H), 0.87 (t,  $J = 7.0$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  
 350  $\text{CDCl}_3/\text{CD}_3\text{OD}$ )  $\delta$  175.8, 104.2, 81.2, 75.8, 74.8, 73.2, 70.5, 70.1, 68.3, 62.3, 51.2, 37.3, 32.8,  
 351 32.6, 30.6, 30.6, 30.4, 30.3, 30.2, 28.0, 26.9, 26.8, 23.5, 14.8; HRMS (TOF)  $m/z$  calcd for  
 352  $\text{C}_{48}\text{H}_{94}\text{NO}_{12}\text{S}$   $[\text{M} - \text{Na}]^+$  908.6502, found 908.6465.

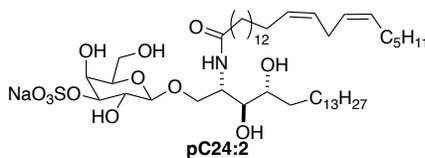
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355  
 356 **(2*S*,3*S*,4*R*)-1-(3-*O*-Sodiumsulfonyl-β- *D*-galactopyranosyloxy)-2-(*N*-15*Z*-tetracosenoyl-**  
 357 **amino)octadecane-3,4-diol (pC24:1).** The general sulfation procedure was followed, and  
 358 sulfatide **pC24:1** was isolated as a white solid (5.3 mg, 58%): mp 211.4–212.4 °C;  $[\alpha]^{25}_D$  8.33 (*c*  
 359 0.50, CHCl<sub>3</sub>/MeOH, 3:2); IR (neat) 3367 (br), 2917, 2850, 1643, 1466, 1224, 1066, 812 cm<sup>-1</sup>; <sup>1</sup>H  
 360 NMR (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 7.78 (m, 1H), 5.37–5.30 (m, 2H), 4.34 (d, *J* = 7.7 Hz,  
 361 1H), 4.29–4.24 (m, 2H) 3.81 (dd, *J* = 11.8, 3.1 Hz, 1H), 3.76–3.56 (m, 2H), 3.70–3.64 (m, 2H),  
 362 3.59–3.57 (m, 2H), 2.20 (t, *J* = 7.6 Hz, 2H), 2.04–2.00 (m, 4H), 1.64–1.50 (m, 4H), 1.44–1.26  
 363 (m, 56H), 0.87 (t, *J* = 6.4 Hz, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD 3:2) δ 175.8, 130.8,  
 364 104.2, 81.3, 75.8, 74.9, 73.2, 70.5, 70.1, 68.3, 62.3, 51.2, 37.4, 37.3, 32.9, 32.8, 30.7, 30.7, 30.6,  
 365 30.6, 30.5, 30.4, 30.4, 30.3, 30.2, 30.2, 28.1, 27.0, 26.9, 23.6, 14.8; HRMS (TOF) *m/z* calcd for  
 366 C<sub>48</sub>H<sub>90</sub>NO<sub>12</sub>S [M – Na]<sup>+</sup> 906.6340, found 906.6339.

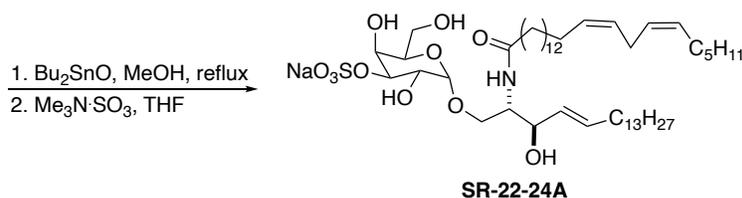
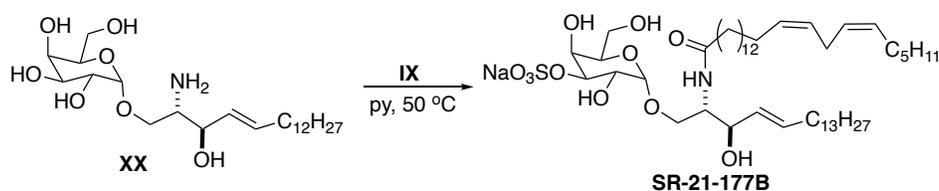
367



368  
 369 **(2*S*,3*S*,4*R*)-1-(3-*O*-Sodiumsulfonyl-β- *D*-galactopyranosyloxy)-2-(*N*-15*Z*,18*Z*-tetraco-**  
 370 **sadienoylamino)octadecan-3,4-diol (pC24:2).** The general sulfation procedure was followed,  
 371 and sulfatide **pC24:2** (containing small amounts of alternative acyl chain isomers) was isolated  
 372 as an off white solid (8.0 mg, 48%): mp 172.0–173.0 °C; IR (neat) 3400 (br), 2917, 2850, 1637,  
 373 1467, 1226, 1061 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD, 3:2) δ 5.41–5.30 (m, 4H), 3.81–  
 374 3.78 (m, 1H), 3.75–3.72 (m, 2H), 3.69–3.64 (m, 2H), 3.60–3.57 (m, 2H), 2.77 (t, *J* = 6.6 Hz,

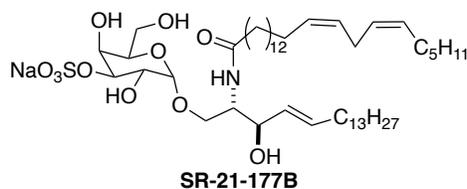
375 1H), 2.20 (t,  $J = 7.3$  Hz, 2H), 2.07–2.02 (m, 4H), 1.59–1.52 (m, 4H), 1.38–1.26 (m, 54H), 0.88  
 376 (m, 6H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3/\text{CD}_3\text{OD}$ , 3:2)  $\delta$  175.8, 131.1, 131.0, 128.9, 128.9, 104.2,  
 377 81.3, 75.8, 74.8, 73.3, 70.5, 70.1, 68.3, 62.3, 54.4, 37.3, 37.3, 32.8, 32.6, 32.4, 30.6, 30.5, 30.4,  
 378 30.3, 30.2, 28.1, 28.1, 26.9, 26.8, 26.5, 23.5, 23.4, 14.8; HRMS (TOF) calcd for  $\text{C}_{48}\text{H}_{92}\text{NO}_{12}\text{S}^-$   
 379  $[\text{M} - \text{Na}]^+ m/z$  904.6189, found 904.6210.

380

381 **Preparation of SR-21-177B and SR-22-24A**

382

383



384

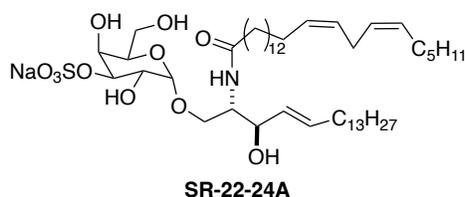
385 **(2S,3S,4E)-1-( $\alpha$ -D-Galactopyranosyloxy)-2-(N-15Z,18Z-tetracosadienoylamino)octadecan-**386 **3,4-diol (SR-21-177B).** *p*-Nitrophenyl 15Z,18Z-tetracosadienoate (**IX**) (36 mg, 0.074 mmol)387 was added to a solution of (2S,3S,4R)-2-amino-1-( $\alpha$ -galactopyranosyloxy)octadecan-3,4-diol388 (**XX**)(6) (31 mg, 0.067 mmol) in pyridine (2 mL). The mixture was stirred in a preheated oil bath

389 at 50 °C for 24 h. The reaction mixture was concentrated and purified by gravity column

390 chromatography on silica gel ( $\text{CH}_2\text{Cl}_2/\text{MeOH}$ , 90:10) to give **SR-21-177B** (26 mg, 48%) as a

391 white solid:  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3/\text{CD}_3\text{OD}$ , 1:1)  $\delta$  7.47 (d,  $J = 8.7$  Hz, 1H), 5.70 (ddd,  $J =$   
 392 14.8, 6.7, 6.7 Hz, 1H), 5.38 (dd,  $J = 15.4$ , 7.0 Hz, 1H), 5.39–5.28 (m, 4H), 4.86, (d  $J = 3.6$  Hz,  
 393 1H), 4.05 (dd,  $J = 6.9$ , 6.9 Hz, 1H), 3.96–3.91 (m, 2H), 3.80–3.72 (m, 6H), 2.76 (t,  $J = 6.4$  Hz,  
 394 2H), 2.18 (t,  $J = 7.6$  Hz, 2H), 2.06–1.99 (m, 6H), 1.58–1.56 (m, 2H), 1.36–1.25 (m, 46H), 0.89–  
 395 0.88 (m, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3/\text{CD}_3\text{OD}$ , 1:1) 174.5, 133.8, 129.7, 129.0, 127.6, 99.7,  
 396 71.8, 70.5, 70.0, 69.5, 68.8, 67.2, 61.4, 53.5, 36.1, 32.1, 31.6, 31.2, 29.4, 29.3, 29.3, 29.2, 29.1,  
 397 28.0, 26.9, 26.8, 25.7, 25.3, 22.3, 22.2 13.5, 13.5; HRMS (ESI) calcd for  $\text{C}_{48}\text{H}_{90}\text{NO}_8$   $[\text{M} + \text{H}]^+$   
 398  $m/z$  808.6667, found 808.6691.

399



400

401 **(2*S*,3*R*,4*E*)-1-(3-*O*-Sodiumsulfonyl- $\alpha$ -*D*-galactopyranosyloxy)-2-(*N*-15*Z*,18*Z*- tetracosa-**

402 **dienylamino)octadec-4-en-3-ol (SR-22-24A).** The general sulfation procedure was followed,

403 and sulfatide **SR-22-24A** was isolated as a white solid (13 mg, 45%):  $^1\text{H}$  NMR (400 MHz,

404  $\text{CDCl}_3/\text{CD}_3\text{OD}$ , 1:1)  $\delta$  7.67 (d,  $J = 8.9$  Hz, 1H), 5.73 (ddd,  $J = 14.8$ , 7.2, 7.2 Hz, 1H), 5.44 (dd,  $J$

405 = 15.4, 7.2 Hz, 1H), 5.39–5.28 (m, 4H), 4.92, (d  $J = 3.8$  Hz, 1H), 4.49 (dd,  $J = 10.2$ , 3.1 Hz, 1H),

406 4.34 (m, 1H), 4.11 (dd,  $J = 7.5$ , 7.5 Hz, 1H), 4.00 (dd,  $J = 10.3$ , 3.8 Hz, 1H), 3.97–3.93 (m, 1H),

407 3.84 (dd,  $J = 5.6$ , 5.6 Hz, 1H), 3.80–3.72 (m, 4H), 2.76 (t,  $J = 6.3$  Hz, 2H), 2.20 (t,  $J = 7.6$  Hz,

408 2H), 2.07–2.00 (m, 6H), 1.58–1.56 (m, 2H), 1.35–1.25 (m, 46H), 0.89–0.86 (m, 6H);  $^{13}\text{C}$  NMR

409 (100 MHz,  $\text{CDCl}_3/\text{CD}_3\text{OD}$ , 1:1)  $\delta$  174.9, 134.1, 129.9, 129.9, 129.4, 127.8, 99.6, 77.771.5, 70.6,

410 68.2, 67.3, 66.9, 61.6, 53.7, 36.3, 32.3, 31.8, 31.4, 29.6, 29.5, 29.4, 29.4, 29.3, 29.2, 29.2, 27.1,

411 27.1, 25.9, 25.4, 22.5, 22.4, 13.7, 13.6; HRMS (ESI) calcd for  $C_{48}H_{88}NNa_2O_{11}S$   $[M + H]^+$   $m/z$   
412 932.5874, found 932.5877.

413

#### 414 **Reagents**

415 Fluorescent protein labeled monoclonal antibodies used for flow cytometry were obtained as  
416 follows: Mouse: anti-CD8 $\alpha$  (clone 53-6.7), anti-CD86 (clone GL-1), anti-CD80 (clone 16-  
417 10A1), anti-CD70 (clone FR70), anti-PDL1 (clone 10F.9G2), anti-PDL2 (clone TY25), anti-  
418 CD45 (clone 30-F11) and anti-CD69 (clone H1.2F3) antibodies were purchased from Biolegend  
419 (San Diego, CA, USA). Anti-B220 (clone RA3-6B2), anti-CD3 (clone 145-2C11), anti-CD1d  
420 (clone 1B1), anti-CD11c (clone HL3), anti-CD45 (clone 30-F11), anti-TCR $\beta$  (clone H57-597),  
421 anti-CD11b (clone M1/70) and anti-CD40 (clone 3/23) antibodies were purchased from BD  
422 BioSciences (San Jose, CA, USA). Anti CD11c (clone REA754) was purchased from Miltenyi  
423 Biotec (Gaithersburg, MD, USA). Anti-CD11b was purchased from eBioscience (San Diego,  
424 CA, USA). PBS57 ( $\alpha$ -GalCer analogue)-loaded CD1d tetramer was obtained from the NIH  
425 Tetramer Core Facility (Emory University, Atlanta, GA, USA). Human: anti-CD3 (clone SP43-  
426 2), anti-IFN $\gamma$  (clone B27) were purchased from BD BioSciences (San Jose, CA, USA), and  
427 LIVE/DEAD Fixable Blue Dead Cell Stain was purchased from Invitrogen (Carlsbad, CA,  
428 USA).

429

430 **Supplementary Fig. Legend**

431

432 **Supplementary Fig. S1. The injection of C24:2 stimulated much more cytokine production**  
433 **in serum than C24:1.**

434 Heat map representing color-coded expression levels of cytokine profiles of mice injected i.p.  
435 with the vehicle used to dissolve the sulfatide analogues, 500 pmol of KRN7000, or 30 nmol of  
436 sulfatide analogues is shown. Serum samples were collected 3 h, 6 h, 12 h, and 24 h after lipid  
437 injection and analyzed. n=5 mice per group. Each row represents an individual mouse.

438

439 **Supplementary Fig. S2. Effects of C24:2 treatment on CD1d surface expression.**

440 BALB/c mice were injected with 30 nmol of C24:2 i.p. CD1d expression of splenocytes was  
441 assessed by flow cytometry. Results are representative data from two experiments (mean  $\pm$  SD)  
442 (n=3 mice per group). (A) Representative flow plot schematic. (B) Quantified MFI of CD1d  
443 expression of various CD45<sup>+</sup> cells.

444

445 **Supplementary Fig. S3. Alpha anomer of C24:2 and C24:1.**

446 (A) Structures of the alpha-anomer of C24:2 (SR-22-24A), the alpha anomer of bGalCer C24:2  
447 (SR-21-177B), and the alpha-anomer of C24:1 ( $\alpha$ C24:1). (B) 50,000 BMDC were incubated  
448 with glycolipid for 3 h and subsequently co-incubated with DN32 cells at a 1:1 ratio overnight.  
449 IL-2 secretion in supernatant from DN32 cells was assessed by ELISA. Results are  
450 representative data from two experiments (mean  $\pm$  SD). (C) CD1d-lipid complexes were adhered  
451 to 96-well plates and co-cultured with DN32 cells overnight. IL-2 secretion in supernatant from

452 DN32 cells was assessed via ELISA. Results are representative data from two experiments  
453 (mean  $\pm$  SD).

454

455 **Supplementary Fig. S4. Representative flow schematic of ICS of human PBMCs.**

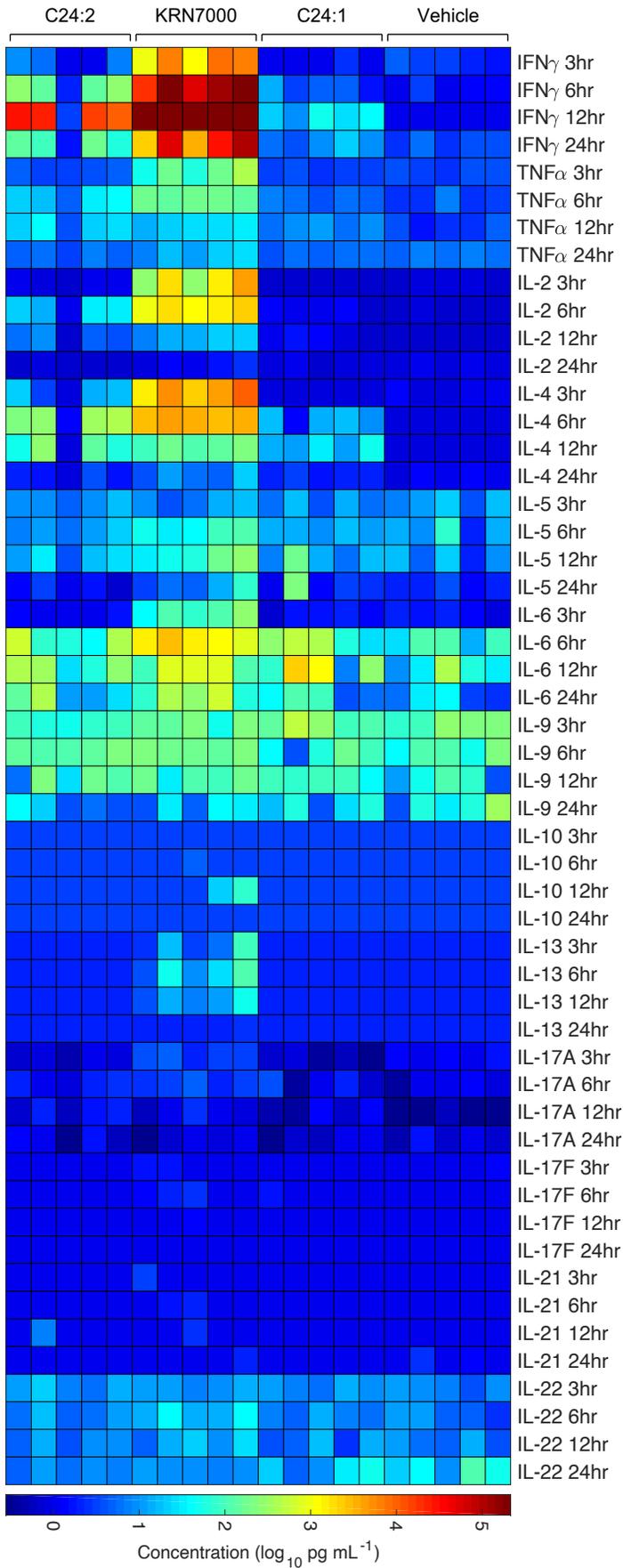
456 Representative flow schematic of  $1 \times 10^6$  healthy human PBMCs were cultured with 10  $\mu\text{g/mL}$   
457 of glycolipid (C24:2 with and without BAF 50 nM) for 15 h then 1 h with brefeldin A.

458 Additionally, human PBMCs were cultured with cell activation cocktail in the presence of BAF.

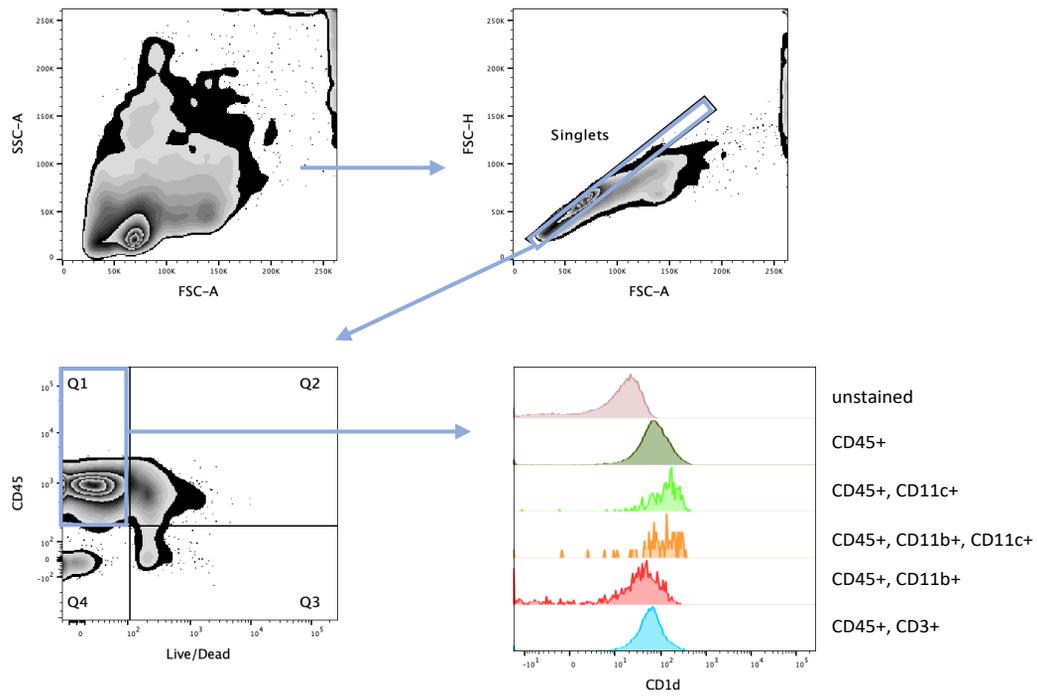
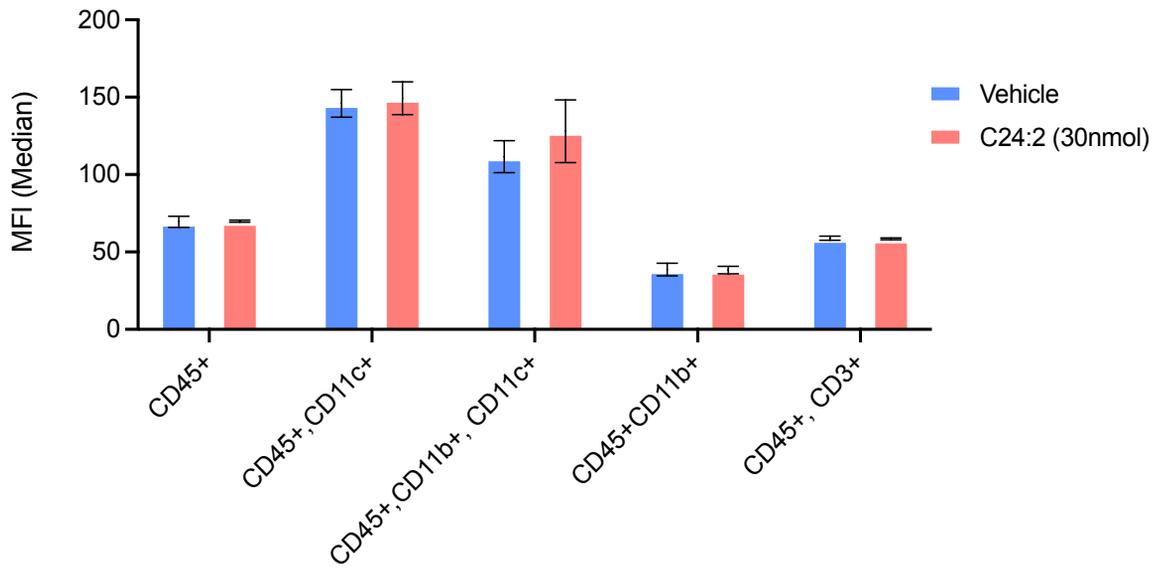
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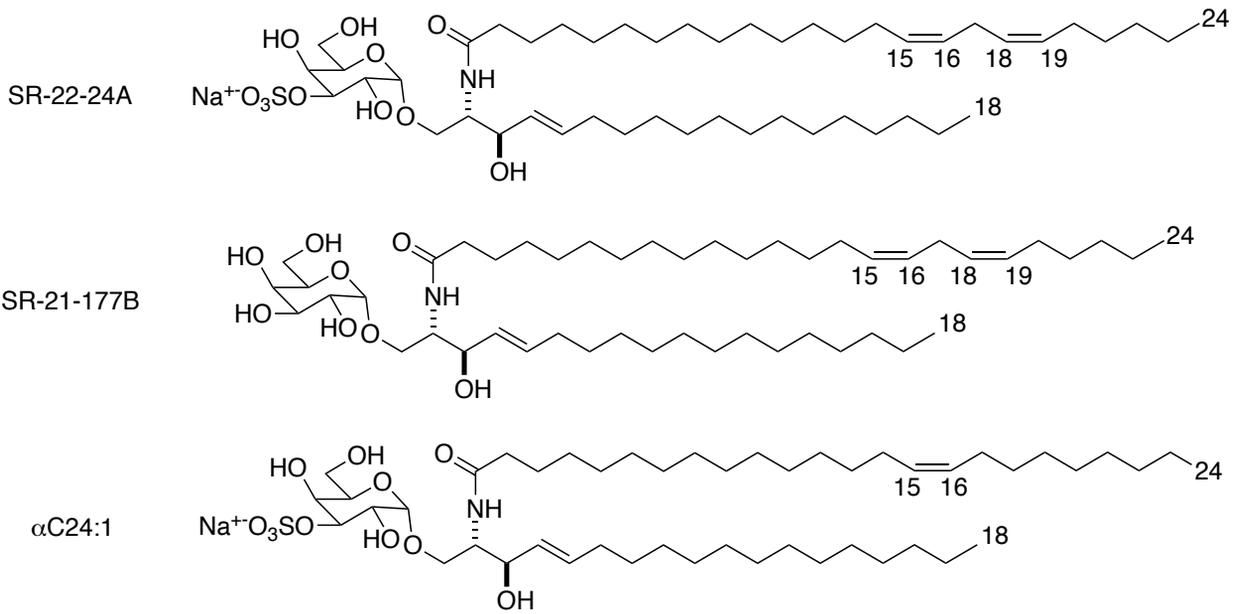
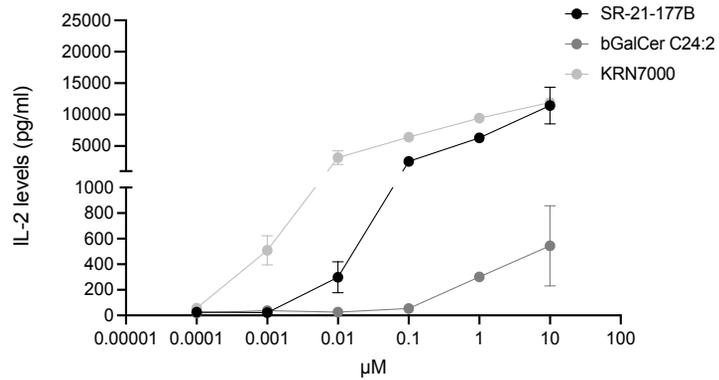
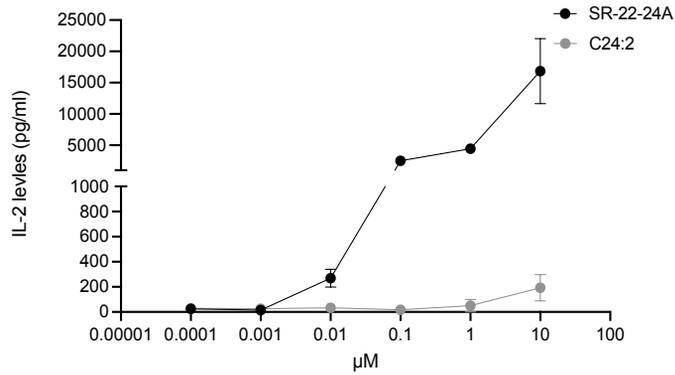
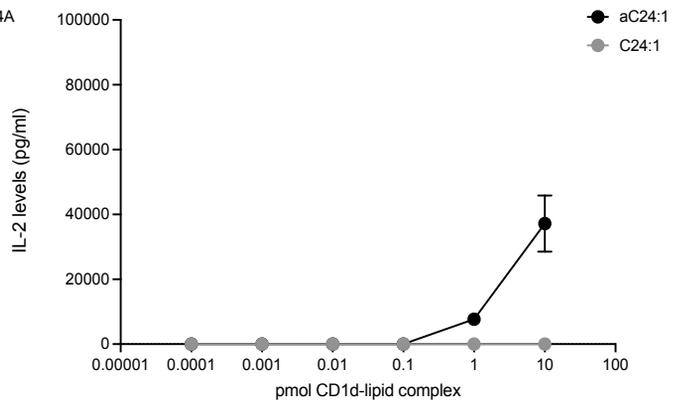
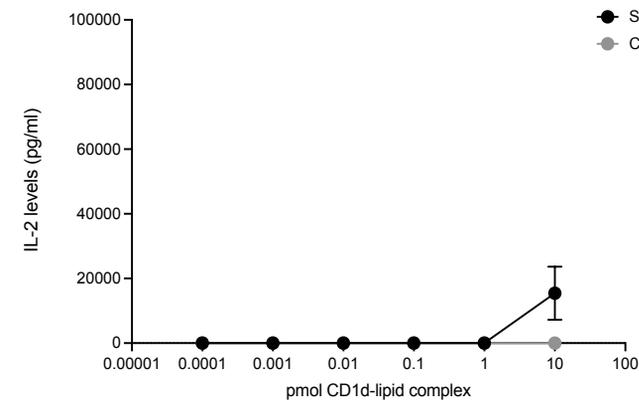
460 **Supplementary References**

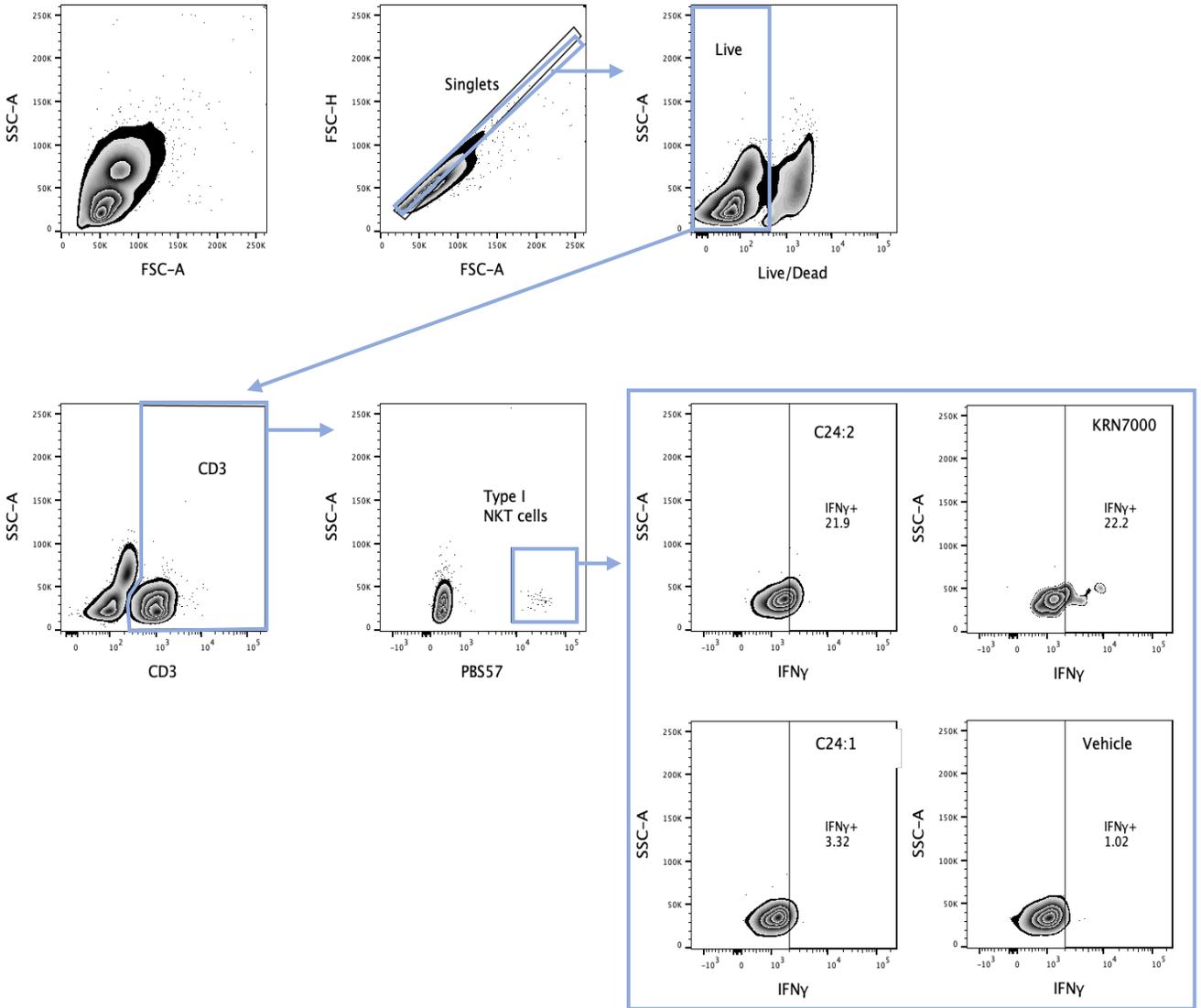
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476



Supplementary Figure S1

**A****B**

**A****B****C**



**Supplementary Figure S4**